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2-chome

Nishikyo-ku, Kyoto 610-11(JP)

Inventor: **Oshika, Yuri**

5-302, Kamikita 5-chome, Hirano-ku

Osaka 547(JP)

Inventor: **Yamada, Takao**

552-2, Hitotsuya-cho, Matsubara

Osaka 580(JP)

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⑯ Applicant: **TAKEDA CHEMICAL INDUSTRIES,
LTD.**

**1-1, Doshomachi 4-chome
Chuo-ku, OSAKA(JP)**

⑯ Representative: **von Kreisler, Alek,**

Dipl.-Chem. et al

Patentanwälte Von Kreisler-Selting-Werner,

Deichmannhaus am Hauptbahnhof

W-5000 Köln 1(DE)

⑯ Inventor: **Fukuda, Tsunehiko
10-202, 9 Oharanonishisakaidani-cho**

⑯ Synthetic gene coding for human parathyroid hormone.

⑯ Disclosed are (1) a DNA containing a synthetic gene for expression of human parathyroid hormone represented by the following DNA sequence:

TCTGTG TCCGAGATTG AGTTAATGCA TAACCTTGGC
 AACACATTGGA ACTCCATGGA GCGTGTAGAA TGGCTGCGTA
 AGAACATTGCA GGATGTGCAC AATTTTGTGTTG CCTTCTGGTGC
 CCCATTGGCT CCTCGTGATG CTGGTTCCCA AAGACCACGT
 AAAAAGGAAG ACAATGTCTT AGTTGAGAGC CATGAAAAAT
 CCCTAGGCGA GGCAGACAAG GCCGATGTGA ATGTATTAAC
 TAAAGCTAAA TCCCCAG

EP 0 483 509 A1

(2) a method for producing the DNA described in (1), which comprises enzymatically ligating a plurality of oligodeoxynucleotides to one another to form the DNA and inserting the DNA into a vector if necessary, (3) a transformant transformed by the DNA described in (1), and (4) a method for producing human parathyroid hormone which comprises cultivating the transformant described in (3), accumulating human parathyroid hormone in a culture medium, and collecting the same, whereby hPTH can be allowed to express in large amounts in a system using E. coli as a host.

BACKGROUND OF THE INVENTION

5 The present invention relates to a recombinant DNA technique for producing human parathyroid hormone (hereinafter referred to as hPTH), and more specifically to a synthetic gene corresponding to the amino acid sequence of hPTH, DNA containing it, a host cell transformed by the DNA, and a method for producing hPTH using the transformant.

10 hPTH is a polypeptide hormone secreted from the parathyroid consisting of 84 amino acids, and one of the most important regulators for calcium metabolism. The clinical application of hPTH to various bone diseases such as hypoparathyroidism and osteoporosis has therefore been strongly desired. However, natural hPTH exists in very small amounts and it is also a protein having a relatively low molecular weight. For this reason, though hPTH can be chemically synthesized, the synthesis thereof requires utilizing techniques which require a high level of skill and which are difficult. Accordingly, the production of hPTH using recombinant DNA techniques is desirable.

15 The DNA sequence of hPTH was first revealed by G. N. Hendy et al. Proc. Natl. Acad. Sci. U.S.A. 78, 7365-7369 (1981). Then, two or three groups attempted to synthesize hPTH using microorganisms to clone cDNA of hPTH and insert it into various expression vectors. Aleström et al. attempted to express and secrete cDNA-derived genes in yeast (Japanese Patent Unexamined Publication No. 2-501108/1990). However, they did not succeed in isolating and identifying hPTH. A. Høgset et al. prepared a gene containing a Met-Gly residue at the N-terminus of hPTH from an hPTH cDNA clone, and expressed the 20 gene in Escherichia coli Biochem. Biophys. Res. Commun. 166, 50-60 (1990). However, the resulting hPTH protein had a Met-Gly residue added to the N-terminus thereof and did not exhibit the activity of hPTH. E. Wingender et al. expressed hPTH in E. coli as a fused protein with β -galactosidase J. Biol. Chem. 246, 4367-4373 (1989). However, this protein whose unnecessary portions were removed was a molecule having Pro added to the N-terminus. A. Høgset et al. succeeded in ligating cDNA of hPTH downstream from a 25 Staphylococcus aureus-protein A promoter and a signal peptide gene, expressing and secreting it outside the cells by E. coli, as well as isolating and identifying hPTH J. Biol. Chem. 265, 7338-7344 (1990). However, the yield is only about 1 mg/liter of culture solution, and the expression amount is insufficient from an industrial viewpoint, particularly when the loss of the product during the purification process is taken into account.

30 When a gene encoding a protein of a certain living organism is intended to be expressed in other hosts, the method of synthesizing that gene chemically is also used as a method for obtaining the gene, in addition to the method of using the gene derived from that living organism. In recent years, automatic synthesis techniques of DNA oligomers have been developed. When the molecular weight of the protein is relatively low and the amino acid sequence or DNA sequence thereof is known, the method of synthesizing 35 the gene is frequently more advantageous for the following reasons:

- (1) The desired gene is relatively easily obtained;
- (2) A terminal sequence used when inserting the gene into a plasmid or a recognition sequence of a restriction enzyme appropriate for modification of the gene can be freely introduced; and
- (3) A gene sequence can be designed so that the desired gene product is highly expressed.

40 W. L. Sung et al. synthesized a structural gene encoding hPTH, using an amino acid codon most acceptable by yeast Biochem. Cell. Biol. 64, 133-138 (1986). Also S. A. Rabban et al. expressed this gene in E. coli, J. Biol. Chem. 263, 1307-1313 (1988), however, its yield of about 200 μ g/l was very low and thus not practical.

45 As described above, attempts have been made to express hPTH in various systems using the cDNA-derived or synthetic genes. In either case, however, the yield was very low and insufficient from an industrial viewpoint.

SUMMARY OF THE INVENTION

50 The present invention provides an amino acid sequence of hPTH, appropriate for solving the above-described problems, as well as a method for producing the gene, recombinant DNA containing the gene, a host cell transformed by the DNA and a method for producing hPTH using the transformant.

In accordance with the present invention, there is provided (1) DNA containing a synthetic gene for expression of human parathyroid hormone represented by the following DNA sequence:

TCTGTG TCCGAGATTC AGTTAATGCA TAACCTTGGC
 AAACATTGGA ACTCCATGGA GCGTGTAGAA TGGCTGCGTA
 5 AGAAGTTGCA GGATGTGCAC AATTTGTTG CCTTAGGTGC
 CCCATTGGCT CCTCGTGATG CTGGTTCCCA AAGACCACGT
 AAAAAGGAAG ACAATGTCTT AGTTGAGAGC CATGAAAAAT
 10 CCCTAGGCGA GGCAGACAAG GCCGATGTGA ATGTATTAAC
 TAAAGCTAAA TCCCAG

15 (2) a method for producing the DNA described in (1), which comprises enzymatically ligating a plurality of oligodeoxynucleotides, one to another, to form the DNA and inserting the DNA into a vector if necessary, (3) a host cell transformed by the DNA described in (1), and (4) a method for producing human parathyroid hormone which comprises cultivating the transformant described in (3), accumulating human parathyroid hormone in a culture medium, and collecting the same.

20 **BRIEF DESCRIPTION OF THE DRAWINGS**

Fig. 1 shows an amino acid sequence and a nucleotide sequence of the synthetic gene of the present invention encoding hPTH;
 25 Fig. 2 is a schematic representation showing an example of cleavage of DNA fragments in synthesizing the hPTH gene of the present invention;
 Fig. 3 is a schematic representation showing an example of DNA fragments for producing synthetic genes of the present invention encoding hPTH;
 Fig. 4-1 is a schematic representation illustrating the production of a synthetic hPTH gene by ligating the 30 respective fragments shown in Fig. 3 one to another;
 Fig. 4-2 is a schematic representation illustrating the production of a synthetic hPTH gene by ligating the respective fragments shown in Fig. 3 one to another;
 Fig. 5 is a schematic representation showing the construction of a plasmid for expression into which a synthetic gene of the present invention encoding hPTH is inserted;
 35 Fig. 6 shows electrophoresis and Western blot results indicating the production of hPTH in a transformant by a gene of the present invention, in which lane 1 shows a molecular weight marker, lane 2 shows a standard sample of hPTH, lane 3 shows an extract (corresponding to 10 μ l of culture solution) of a transformant induced by isopropyl- β -D-thiogalactopyranoside (IPTG), and lane 4 shows an extract (corresponding to 10 μ l of culture solution) of a transformant not induced by IPTG;
 40 Fig. 7 shows electrophoresis and Western blot results indicating the production of hPTH in a transformant by a gene of the present invention, in which lane 1 shows a prestained molecular weight marker, lane 2 shows an extract (corresponding to 10 μ l of culture solution) of a transformant not induced by IPTG, lane 3 shows an extract (corresponding to 10 μ l of culture solution) of a transformant induced by IPTG, and lane 4 shows a standard PTH sample;
 45 Fig. 8 shows an elution pattern of purified hPTH obtained by reversed phase high performance liquid chromatography;
 Fig. 9 shows electrophoresis and Western blot results for an hPTH sample, in which lane 1 shows a prestained molecular weight marker, lane 2 shows purified hPTH obtained by the present invention, and lane 3 shows a standard hPTH sample;
 50 Fig. 10 shows an electrophoresis and Western blot results for an hPTH sample, in which lane 1 shows a prestained molecular weight marker, lane 2 shows purified hPTH obtained by the present invention, and lane 3 shows a standard hPTH sample; and
 Fig. 11 is a graph showing the biological activity of hPTH fragments, in which - - shows a synthetic hPTH(1-34) fragment, and --- shows purified hPTH obtained by the present invention.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

Sequences of synthetic genes are designed so as to satisfy various conditions to enhance the expression of the genes. One of the conditions is that codons most acceptable by cells of expression systems should be employed. Accordingly, in the hPTH genes of the present invention, codons are employed which are frequently used for the production of proteins by recombinant technology and accepted by both E. coli and yeast. Further, consideration is given to arranging recognition sequences of restriction enzymes convenient for modification of the genes at appropriate positions when selecting the codons. Furthermore, to construct the desired genes accurately, relatively long, unnecessary palindrome sequences and directly repeated sequences existing on DNA chains are removed as much as possible. As additional conditions to enhance the expression of the genes, the stability and translation efficiency of mRNAs corresponding to structural genes should be considered. In this case, secondary structure determined by nucleotide sequences of mRNAs is believed to be of importance. Therefore, for the hPTH genes of the present invention, hairpin structures and the like are searched for using a computer during the course of design, and the sequence is then modified, preferably by removing it. Thus, a novel DNA sequence most suitable for the production of hPTH as shown in Fig. 1 (SEQ ID NO:1) has been discovered. This gene sequence is about 25.4% different from the above-described gene of W. L. Sung et al. in DNA sequence. In addition to the DNA sequence, the amino acid sequence is shown in Fig. 1.

The gene can be expressed as either a fused peptide or hPTH directly without forming the fused peptide.

In the former case, either a DNA sequence starting with an initiation codon ATG and coding for a protein other than hPTH may be arranged at the 5'-terminal side of the synthetic gene of hPTH, and it ends in a stop codon (for example, TAA); or a DNA sequence coding for a protein other than hPTH may be arranged at the 3'-terminal side of the synthetic gene of hPTH starting with an initiation codon ATG and it ends in a stop codon (for example, TAA).

In the latter direct expression, the initiation codon ATG and a stop codon (for example, TAA) are directly arranged on the 5'- and 3'-terminal sides, respectively, and the 5'- and 3'-termini have, for example, NdeI and BamHI cohesive ends, respectively, for insertion into a vector, in addition to a sequence coding for a polypeptide of hPTH, as shown in Fig. 2 (SEQ ID NO:2).

In synthesizing the hPTH gene of the present invention, the hPTH gene is finally cleaved into 14 fragments, for example, as shown in Fig. 2 (SEQ ID NO:2, SEQ ID NO:3). In this case, care is taken so as to avoid self-linkage between fragments, e.g. a self-complementary sequence at the 5'- or 3'-terminus. The respective fragments are shown in Fig. 3. Methods for cleaving the gene into the fragments are not limited to the above-mentioned method, and various methods are available as long as the above self-linkage is avoidable.

Fragments #1 to #14 (SEQ ID NOS: 4 to 17) can be produced by known synthesizing methods. For the fragments, except for #1 and #14, the termini are phosphorylated at 5'-termini with polynucleotide kinase as so required, and all of the fragments are hybridized to ligate them to one another with DNA ligase, or the phosphorylated fragments first divided into two or three groups are hybridized to form a double stranded DNA fragment with DNA ligase, and the DNA fragments of the respective groups are further ligated to one another with DNA ligase, thereby obtaining a complete double stranded hPTH gene (refer to Fig. 4).

The resulting gene is ligated to a product obtained by digesting pUC19 with NdeI and BamHI to obtain a novel plasmid pU⁺PTH⁺C19, and E. coli JM109 is transformed. As to the isolated plasmid, the nucleotide sequence is determined by the Sanger method using a portion of the DNA fragment as a primer. More easily, the DNA fragment obtained by NdeI-BamHI digestion is digested with AvrII, Ncol, HgiAI or Alul to give a correct restriction site, thereby confirming the existence of the desired hPTH gene.

When the synthetic gene of the present invention is expressed, it is preferably used as a recombinant DNA inserted into vectors such as plasmids or bacteriophages.

Examples of such plasmids include pBR322 [Gene 2, 95 (1977)], pBR325 [Gene 4, 121 (1978)], pUC12 [Gene 19, 259 (1982)] and pUC13 [Gene 19, 259 (1982)], each derived from Escherichia coli, and pUB110 derived from Bacillus subtilis [Biochemical and Biophysical Research Communication 112, 678 (1983)]. However, any other plasmids can be used as long as they are viable in the host cells. Examples of the phage vectors include λgt11 R. Young and R. Davis, Proc. Natl. Acad. Sci. U.S.A. 80, 1194 (1983). However, any other phag vectors can be used as long as they are viable in the host cells.

The plasmids thus obtained are introduced into appropriate host cells such as Escherichia, Bacillus, yeast and animal cells.

Methods for transforming the host cells with the plasmids include, for example, the calcium chloride method or the calcium chloride/rubidium chloride method described in T. Maniatis et al., Molecular Cloning, Cold Spring Harbor Laboratory, p.249 (1982).

When phage vectors are used, for example, they can be transduced into proliferated E. coli, using the in vitro packaging method.

It is preferred that the above-mentioned recombinant DNA has a promoter upstream from the above-mentioned initiation codon ATG, and as the promoter, any promoter may be employed as long as it is suitable for the host cells used for the production of transformants.

For example, a trp promoter, a lac promoter, a recA promoter, a λPL promoter, a lpp promoter, T7 promoter, etc. may be used for E. coli (for example, MM294, BL21, BL21/PLys, W3110, DH1 and N4830); a SP01 promoter, a SP02 promoter, a penP promoter, etc. may be used for Bacillus subtilis (for example, MI114); a promoter for major cell wall proteins, etc. may be used for Bacillus brevis; a PH05 promoter, a PGK promoter, a GAP promoter, an ADH promoter, etc. may be used for Saccharomyces cerevisiae (for example, AH22); and an SV40-derived promoter, mouse leukemia virus (MuLV)-derived LTR region promoter, etc. may be used for animal cells (for example, monkey cell COS-7 and Chinese hamster ovary cell (CHO)). In particular, it is preferred that the promoter is the T7 promoter, the trp promoter or the λPL promoter when the host cell is E. coli, and that an enhancer is used in addition to the above-mentioned promoter when the host cell is an animal cell.

The transformants thus obtained are cultivated in the known media. The media for the transformants of Escherichia coli include, for example, LB medium.

The transformants of Escherichia coli are cultivated usually at 15 to 43°C for 2 to 24 hours, preferably at 28 to 40°C for 4 to 16 hours, with aeration or agitation if necessary. After cultivation, the cells are collected by methods known in the art. In the case of the transformants of Escherichia coli, the cells are suspended in buffer solutions or buffer solutions containing protein denaturants such as urea and guanidine hydrochloride, and disrupted by mechanical disruption such as ultrasonic treatment, lysozyme treatment and glass bead treatment, followed by centrifugation to obtain extracted solutions containing hPTH. hPTH can be separated and purified from the extracted solutions by known means. The separating and purifying means include column chromatography such as gel filtration, ion exchange chromatography using a cation exchange resin or an anion exchange resin, hydrophobic chromatography and partition adsorption chromatography, and high performance liquid chromatography. The cultivation of the transformants of the Bacillus, the yeast and the animal cells and the separation and purification of hPTH from the cultures are carried out by methods known in the art.

hPTH thus obtained can be used as a therapeutic agent for various diseases caused by an abnormality of the calcium metabolism, for example, osteoporosis, hypoparathyroidism and hypertension. The dosage thereof is properly determined, taking into account the object of administration, the disease, etc. in each case, and a suitable amount is given within the range of 1 ng to 100 μg/kg of weight a day. Usually, hPTH is given parenterally in combination with pharmaceutically acceptable carriers, excipients or diluents as injections, nasotracheal agents, perrectum agents, transvaginal agents or percutaneous agents, but it may be given orally in some cases.

One example of the expression of the hPTH gene is described below (refer to Fig. 5).

A DNA fragment of 267 base pairs cut out of pU*PTH*C19 with NdeI-BamHI was inserted into a NdeI-BamHI site to obtain a vector pET*PTH*3C for expression under T7 promoter control.

E. coli BL21 was transformed using pET*PTH*3C, and growing colonies were selected based upon ampicillin resistance to obtain a strain containing a desired hPTH gene. This transformant strain was cultivated, and cells were collected by centrifugation. hPTH contained in a solution treated with 7 M guanidine was stained with Coomassie Brilliant Blue (CBB) after SDS-polyacrylamide electrophoresis (SDS-PAGE), or immunoblotting was conducted by an hPTH antibody after SDS-PAGE, thereby determining hPTH. As a result, the product was about 200 mg/l or more (refer to Figs. 6 and 7). This expression amount is epoch-making as the direct expression of hPTH in E. coli.

In the present invention, the codon most acceptable in cells of expression systems is employed as the DNA sequence of the hPTH gene, and the gene is designed, also taking into account the secondary structure of mRNA corresponding to the DNA sequence, whereby the translation efficiency of mRNA are improved, which results in the efficient production of hPTH. Further, in the present invention, the arrangement of the recognition sites of several kinds of restriction enzymes in the structural gene makes the insertion of the gene and the confirmation of an inserting direction easy, and makes the modification of the gene easy when analogue proteins of hPTH will be synthesized in future. Moreover, the gene can be inserted into several kinds of vectors. Thus, the advantage and the diversity on gene manipulation can be exhibited.

Example 11. Synthesis of DNA Fragments

5 Fourteen DNA fragments #1 to #14 (SEQ ID NO: Nos. 4 to 17) shown in Fig. 3 were synthesized using properly protected DNA β -cyanoethylphosphoamidite, and using an automatic synthesizer (Model 380A, Applied Biosystems). As a protocol for synthesis, one specified by Applied Biosystems was used. The protected DNA oligomer-resins thus synthesized (0.2 μ mole of the resin) were heated in 2 ml of concentrated aqueous ammonia at 60°C for 6 hours. The resulting products were purified by reversed
 10 phase high performance liquid chromatography (hereinafter referred to as HPLC) to obtain DNA oligomers only the 5'-terminal hydroxyl groups of which were protected by dimethoxytrityl groups. These DNA oligomers were treated with 2 ml of 80% acetic acid for 20 minutes to remove the terminal dimethoxytrityl groups, and the resulting products were purified by reversed phase HPLC and ion exchange HPLC. The fourteen DNA oligomers thus synthesized are as shown in Fig. 3 (SEQ. NOS: 4 to 17).

15

2. Phosphorylation of DNA oligomers

Each of the twelve DNA oligomers #2 to #13 (SEQ ID NOS: 5 to 16) except for #1 (SEQ ID NO: 4) and #14 (SEQ ID NO: 17) which were to form the 5'-termini was reacted in 25 μ l of a phosphorylation reaction
 20 solution [10 μ l of the DNA oligomer, 50 mM Tris-HCl (pH 7.6), 10 mM MgCl₂, 1 mM spermidine, 10 mM dithiothreitol (hereinafter referred to as DTT), 0.1 mg/ml bovine serum albumin (hereinafter referred to as BSA), 1 mM ATP, 10 units of T4 polynucleotide kinase (Takara Shuzo)] at 37°C for 1 hour to phosphorylate the 5'-terminus. This reaction solution was treated at 65°C for 10 minutes, followed by freezing and thawing. The resulting product was subjected to the following reaction.

25

3. Ligation of DNA Fragments (Refer to Figs. 4-1 and 4-2)

3-1 A series of stages for forming a double stranded structure of an hPTH gene are as shown in Fig. 4-1. Referring to Fig. 4-1, the mark \leftarrow indicates that a 5'-terminal hydroxyl group is phosphorylated. For example, the ligation of block I was carried out as follows. 7.5 μ l portions of the phosphorylated reaction solutions of the five DNA fragments [corresponding to DNA fragments #2 to #6 (SEQ ID NOS: 5 to 9)] obtained by the operation described in the above item 2 were combined with 2.5 μ g of DNA fragment #1 (SEQ NO: 4) corresponding to the 5'-terminus to 50 μ l. Then, 5 units of T4 DNA ligase (New England Biolabs) was added thereto, followed by incubation at 14°C for 5 hours. The resulting product was thereafter treated at 65°C for 10 minutes to terminate the reaction, thus obtaining block I. Blocks II and III were similarly prepared. 20 μ l portions of these block I to III were mixed and 5 units of T4 DNA ligase was added thereto, followed by incubation at 14°C for 20 hours. The resulting product was treated at 65°C for 10 minutes to terminate the reaction.

The product thus obtained was subjected to electrophoresis on a 7.5% polyacrylamide gel in a buffer (pH 8.3, 100 mM Tris-HCl, 100 mM borate, 2 mM EDTA) at 160 V for 1.5 hours. After electrophoresis, the gel was stained with 0.6 mg/l ethidium bromide (EtBr). Gel fragments containing 263-bp DNA fragments were sealed in a dialysis tube and submerged in a buffer for electrophoresis. Then, the DNA fragments were electrically eluted from the gel [J. Mol. Biol. 110, 119 (1977)]. A solution in this dialysis tube was recovered and poured on an Elutip-d column (Schleicher & Schnell) previously buffered with a solution containing 0.2 M NaCl, 20 mM Tris-HCl (pH 7.4) and 1.0 mM EDTA to allow the DNA fragment to be adsorbed. Then, the DNA fragments were eluted with a solution containing 1.0 M NaCl, 20 mM Tris-HCl (pH 7.4) and 1.0 mM EDTA. Twice as much ethanol as the eluate was added to the eluate, and the mixture was cooled to -20°C. Then, the DNA fragments were precipitated by centrifugation.

3-2 A series of stages for forming a double stranded structure of an hPTH gene can also be achieved by a process shown in Fig. 4-2. Referring to Fig. 4-2, the mark \leftarrow indicates that a 5'-terminal hydroxyl group is phosphorylated. 5 μ l portions of the phosphorylated reaction solutions of the twelve kinds of DNA fragments [corresponding to DNA fragments #2 to #13 (SEQ NOS: 5 to 16)] obtained in the above item 2 were combined with 2 μ g of DNA fragments #1 (SEQ ID NO: 4) and #14 (SEQ ID NO: 17) corresponding to the 5'-terminus to 50 μ l. Then, 5 units of T4 DNA ligase (Takara Shuzo) was added thereto, followed by incubation at 15°C for 20 hours.

The product thus obtained was subjected to electrophoresis on a 8% polyacrylamide gel in a buffer (pH 8.3, 100 mM Tris-HCl, 100 mM boric acid, 2 mM EDTA) at 125 V for 2 hours. After electrophoresis, the gel was stained with 0.6 mg/l EtBr. Gel fragments containing 263-bp DNA fragments were sealed in a dialysis

tube and submerged in a buffer for electrophoresis. Then, the DNA fragments were electrically eluted from the gel. A solution in this dialysis tube was subjected to phenol treatment twice, followed by recovery of an aqueous layer (an upper layer). Then, twice as much ethanol as the aqueous layer was added thereto, and the mixture was cooled to -70°C. The DNA fragments were thereafter precipitated by centrifugation. Thus, 5 about 1 µg of the DNA fragments was obtained. After phosphorylation with T4 polynucleotide kinase (Takara Shuzo), the DNA fragments were subjected to the following experiment 4-2.

4. Cloning of hPTH Gene (Fig. 5)

10 4-1 As a cloning vector, *E. coli* plasmid pBR322-derived pUC19 [J. Messing, Gene 33 103-109 (1985)] was used. pUC19 DNA was reacted in 20 µl of a reaction solution [20 mM Tris-HCl (pH 7.6), 7 mM MgCl₂, 150 mM NaCl, 10 mM 2-mercaptoethanol, 20 units of NdeI (New England Biolabs), 15 units of BamHI (Takara Shuzo)] at 37°C for 24 hours. Then, the resulting product was diluted 5 times with water, and treated at 65°C for 20 minutes to inactivate the enzyme. 5 µl of this reaction solution was mixed with about 5 equivalents of the DNA fragments obtained in the above item 3-1 to prepare 20 µl of a reaction solution containing 50 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, 10 mM DTT, 1 mM spermidine, 0.1 mg/ml BSA and 1 mM ATP. Then, T4 DNA ligase (New England Biolab) was reacted with this solution at 14°C for 15 hours to ligate the hPTH gene to the plasmid.

20 Using this reaction solution, the *E. coli* JM109 strain [J. Messing, Gene 33, 103-119 (1985)] was transformed according to methods known in the art. Namely, 50 µl of competent cells [D. Hanahan, J. Mol. Biol. 166, 557 (1983)] stored at -70°C was incubated at 0°C for 15 minutes, and then 10 µl of the above-mentioned reaction solution was added thereto. The resulting solution was further incubated at 0°C for 15 minutes, and then incubated at 42°C for 1.5 minutes and further at 0°C for 2 minutes. To this reaction solution was added 200 µl of LB medium (containing 10 g of Bacto-tryptone, 5 g of a Bacto-yeast extract and 5 g of NaCl), and incubated at 37°C for 1 hour. This *E. coli* was seeded onto LB agar medium containing 50 µg/ml ampicillin, 100 µg/ml X-Gal and 0.1 mM isopropyl-β-D-thiogalactopyranoside (IPTG), and incubated at 37°C overnight. Of the resulting ampicillin-resistant colonies, 14 β-galactosidase-deficient strains were selected and plasmid DNAs of transformed strains thereof were crudely purified by the alkali method [T. Maniatis et al., Molecular Cloning, (Cold Spring Harbor Laboratory) 368-369 (1982)], followed by 25 digestion with NcoI and BamHI, and further with NdeI and BamHI. The electrophoresis patterns of these 30 digests on a 1.7% agarose gel revealed that one strain was a transformed strain into which the hPTH gene was correctly inserted.

35 4-2 The hPTH gene was also cloned by the following method. As a cloning vector, pUC19 (Takara Shuzo) was used. 0.5 µg of pUC19 DNA was reacted in 10 µl of a reaction solution [50 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, 100 mM NaCl, 1 mM dithiothreitol, 20 units of NdeI (New England Biolabs), 10 units of BamHI (Takara Shuzo)] at 37°C for 5 hours. Then, the resulting product was treated at 65°C for 15 minutes to inactivate the enzyme. 1 µl of this reaction solution was mixed with about 10 equivalents of the DNA 40 fragment obtained in the above item 3-2, and the hPTH gene was ligated to the plasmid using a DNA ligation kit (Takara Shuzo). The transformation into the *E. coli* JM109 strain was carried out in the same manner as with the above item 4-1. Of the resulting ampicillin-resistant colonies, 17 β-galactosidase-deficient strains were selected and plasmid DNAs of transformed strains thereof were crudely purified by the alkali method, followed-by-digestion-with-NcoI-and-BamHI, and further with NdeI and BamHI. The 45 electrophoresis patterns of these digests on a 1.5% agarose gel revealed that three strains were transformed strains into which the hPTH gene was correctly inserted.

50 The cloning vectors obtained in the above items 4-1 and 4-2 were named pU*PTH*C19. 20 ml of LB medium containing 50 µg/ml ampicillin was inoculated with one loopful of *E. coli* JM109 recombinants having this plasmid pU*PTH*C19, and cultivated at 37°C overnight with shaking. Plasmid DNA was crudely purified from this culture solution, and dissolved in 80 µl of TE buffer [10 mM Tris-HCl (pH 8.0), 1 mM EDTA] containing 20 µg/ml RNase.

Example 2

Construction of Plasmid for Expression of hPTH and Production of Transformant (Fig. 5)

55 1) About 10 µg of pU*PTH*C19 obtained in the above item 4 of Example 1 was reacted in a reaction solution [150 mM NaCl, 20 mM Tris-HCl (pH 7.8), 7 mM MgCl₂, 10 mM mercaptoethanol, 40 units of NdeI, 20 units of BamHI (Takara Shuzo)] at 37°C for 5 hours. Then, 263-bp DNA fragments were purified by 1.7% agarose gel electrophoresis according to known methods. On the other hand, as a

vector for expression, pET3C [F. W. Stadier et al., *Methods in Enzymology* 195, 60-89 (1990)] was used. pET3C DNA was digested with NdeI and BamHI in the same manner as above, and four times as much water as the resulting reaction solution was added thereto, followed by heating at 65°C for 20 minutes to inactivate the enzymes.

5 Each of the 263-bp DNA and the plasmid DNA has single stranded cohesive ends produced by NdeI digestion and BamHI digestion at both ends thereof.

Both of them were mixed with each other, and the mixture was reacted with T4 DNA ligase (New England Biolabs) in the presence of 50 mM Tris-HCl (pH 7.6), 10 mM MgCl₂, 10 mM DTT, 1 mM spermidine, 0.1 mg/ml BSA and 1 mM ATP at 14°C for 16 hours to ligate the DNAs to each other, followed by transformation of the *E. coli* JM109 strain in the same manner as above. Then, this *E. coli* was seeded onto LB agar medium containing 50 µg/ml ampicillin, and cultivated at 37°C for 1 day. The resulting ampicillin-resistant colonies were selected. Plasmid DNAs of the transformed strains were further digested by combinations of restriction enzymes such as NdeI-BamHI, BglII-BamHI, EcoRI-NdeI and AvrII-BglII. Transformed strains containing the correct hPTH genes were selected by their patterns of polyacrylamide electrophoresis. The plasmids for expression thus obtained were named pE-PTH, and the transformed strains were named *E. coli* JM109/pE-PTH.

2) Plasmid DNAs were isolated from JM109/pE-PTH obtained in the above item 1), and crudely purified to transform *E. coli* MM294(DE3) having T7 RNA polymerase necessary for expression of a T7 promoter (European Patent Unexamined Laid-Open No. 416505). First, 10 ml of LD medium was inoculated with one loopful of *E. coli* MM294(DE3), and cultivated at 37°C with shaking to a Klett of 60 to 180. To this culture solution were added 10% w/v polyethylene glycol, 5% v/v dimethyl sulfoxide and 50 mM MgCl₂ (pH 6.5), and a reaction solution was brought up to 100 µl with the addition of LB medium. 10 mg of the plasmid DNAs was added thereto, and incubated at 4°C for 10 minutes, followed by seeding onto LB agar medium containing 50 µg/ml ampicillin. Then, cultivation was carried out overnight at 37°C.

25 Plasmid DNAs obtained from the resulting colonies similarly with the method described above were digested with restriction enzymes, and transformed strains containing the hPTH genes were selected by their patterns of electrophoresis. These strains were named *E. coli* MM294(DE3)/pE-PTH.

30 *E. coli* MM294(DE3) pE-PTH was deposited with the Institute for Fermentation, Osaka, Japan (IFO) under the accession number IFO 15087 on August 28, 1990, and also deposited with the Fermentation Research Institute, the Agency of Industrial Science and Technology, the Ministry of International Trade and Industry, Japan (FRI) under the accession number FERM BP-3110 on September 25, 1990.

Example 3

35 Production of hPTH

1) *E. coli* MM294(DE3) pE-PTH was cultivated overnight at 37°C in LB medium containing 50 µg/ml ampicillin with shaking. 100 µl of the resulting culture solution was added to 10 ml of the same medium dispensed in 200 ml flasks, and cultivated at 37°C to a Klett of about 170. Then, IPTG was added thereto to a concentration of 0.1 mM. After cultivation was further continued for 2 hours, 1 ml of the culture solution was centrifuged at 15,000 rpm at 4°C for 5 minutes. The resulting cells were dissolved in 100 µl of a solution containing 0.5 M Tris-HCl (pH 6.8), 10% glycerol, 10% (w/v) sodium dodecyl sulfate (SDS), 0.1% (w/v) β-mercaptoethanol and bromophenol blue [U. K. Laemmli, *Nature* 227, 680 (1970)]. After boiling for 3 minutes, the solution was subjected to 16% SDS-polyacrylamide gel electrophoresis (PAGE). After electrophoresis, the gel was stained with Coomassie Brilliant Blue. As a result, an intense band indicating the same mobility as that of a standard hPTH sample was observed (refer to Fig. 6). Further, another gel was subjected to western blotting using an hPTH antibody. As a result, the same stain pattern as with the standard hPTH sample was obtained (refer to Fig. 7). Quantitative comparison with the standard sample for the stain of the gels and western blotting revealed that hPTH is expressed in an amount of about 200 mg per liter of culture solution.

2) hPTH accumulated in *E. coli* was purified in the following manner. Cells from 200 ml of a culture solution obtained similarly with the above-mentioned method were suspended in a buffer (5 ml) containing 8 M urea, 50 mM Tris-HCl (pH 7.5), 50 mM EDTA and 1 mM α-toluenesulfonyl fluoride, and vigorously stirred under ice cooling for about 1 hour to disrupt the cells, followed by centrifugation at 15,000 rpm at 4°C for 20 minutes. The supernatant was recovered, and the precipitate was similarly extracted twice with 3 ml portions of a buffer having the same composition. The extracts were combined with the supernatant, followed by two-fold dilution. The resulting solution was passed through a CM-Toyo Pearl column (10 ml) of TSK-Gel (Tosoh) equilibrated with 50 mM ammonium acetate buffer (pH 5)

containing 4 M urea to allow a desired product to be adsorbed. The column was washed with 50 mM ammonium acetate buffer (pH 5) containing 4 M urea. About 10 ml of the buffer was required for washing. When absorption at 280 nm disappeared, the column was developed by a linear gradient of 50 ml of 50 mM ammonium acetate buffer (pH 5)-50 ml of 0.5 M ammonium acetate buffer (pH 6) (flow rate: 5 ml/hour, volume of 1 fraction: 2 ml). Fractions 33 to 47 were collected and lyophilized. These fractions were subjected to reversed phase HPLC under the following conditions:

Column: YMC-Pack A-325 S-5 120A ODS (1X30 cm) (Y. M. C.)

Solvent: A linear gradient of 25% to 50% acetonitrile containing 0.1% trifluoroacetic acid (for 30 minutes)

Flow rate: 3 ml/minute

The main peak (retention time: 17.0 minutes) contained the desired product. The resulting eluate was passed through a Bio-Rad AG1X8 (acetate form) column (Bio-Rad Laboratory) and the washings were also combined therewith. Then, acetonitrile was removed by distillation, followed by lyophilization. 4.2 mg of desired hPTH was obtained as a white powder.

The following analytical results revealed that this sample was hPTH of high purity.

a) Reversed phase HPLC showed a single sharp peak (refer to Fig. 8).

Column: YMC-Pack A-324 S-5 DDS 120A (10X300 mm)

Eluents: Solution A (0.1% trifluoroacetic acid)

Solution B (acetonitrile containing 0.1% trifluoroacetic acid)

Linear gradient: 0 to 3 minutes (0%, B), 3 to 10 minutes (0 to 30%, B), 10 to 70 minutes (30 to 45%, B)

b) SDS-PAGE and western blotting after electrophoresis also showed a single band of the same mobility as that of the standard sample. The conditions were the same as with Figs. 6 and 7 (refer to Figs. 9 and 10).

c) Amino acid analysis (hydrolyzed with 5.7 N hydrochloric acid in the presence of thioglycollic acid in a sealed tube under reduced pressure at 110 °C for 24 hours, and values in parentheses indicate theoretical values):

Asp (10) 10.5; Thr (1) 0.96; Ser (7) 6.04; Glu (11) 11.73; Pro (3) 2.88; Gly (4) 4.69; Ala (7) 7.03; Val (8) 7.91; Met (2) 1.96; Ile (1) 1.06; Leu (10) 10.6; Phe (1) 1.10; Lys (9) 9.32; His (4) 4.04; Arg (5) 4.00; Trp (1) 0.97

(Recovery: 82.3%)

d) N-terminal amino acid sequence analysis by a gas-phase sequencer Model 470A (Applied Biosystems) revealed that the sequence from the 1-position Ser to the 15-position Leu was correct.

e) In first atom bombardment mass spectrometry analysis [JMS-HX110 (Nihon Denshi) was used] after cyanogen bromide cleavage in 0.1 N hydrochloric acid, the following peaks were observed:

(The 1-position to the 7-position) - homoserine (MH⁺)

Found: 858.5

Theoretical: 858.5

(The 1-position to the 17-position) - homoserine (MH⁺)

Found: 1990.2

Theoretical: 1990.0

(The 9-position to the 17-position) - homoserine (MH⁺)

Found: 1102.7

Theoretical: 1102.7

No peak derived from formylhomoserine or homoserine was observed.

Also in analysis after digestion with asparagine-specific endopeptidase, the following peaks were observed:

(The 1-position to the 16-position) (MH⁺)

Found: 1819.9

Theoretical: 1819.9

(The 17-position to the 33-position) (MH⁺)

Found: 2168.8

Theoretical: 2168.1

(The 58-position to the 76-position) (M + Na)

Found: 2061.8

Theoretical: 2062.0

(The 77-position to the 84-position) (MH⁺)

Found: 874.4

Theoretical: 874.5

From the results obtained above, it was judged that hPTH obtained here did not contain Met or formyl-

5 Met at the N-terminus thereof and had a structure starting with the 1-position Ser and ending in the C-terminal 84-position Gln.

Experimental Example

10 The biological activity of hPTH thus obtained was assayed by a modified method of Shigeno et al. [The Journal of Biological Chemistry 263, 18369-18377 (1988)]. A culture solution (Hank's solution containing 20 mM N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid (HEPES), 0.1% bovine serum albumin and 0.5 mM isobutylmethylxanthine) containing 0.01, 0.1, 1, 10 or 100 nM the present analogue was added in an amount of 100 μ l to a mouse cranial bone-derived osteoblast-like cell strain, MC3T3-E1 cells, cultivated on a 15 96-well multiplate (Nunclon, Nunc), followed by reaction at room temperature for 30 minutes. After addition of 100 μ l of 0.2 N hydrochloric acid, the mixture was immersed in boiling water for 2.5 minutes, and cyclic adenosine monophosphate (cAMP) produced by a PTH receptor was extracted from the cells. The total cAMP in the culture solution and the cells was assayed using a commercial radioimmunoassay kit (cyclic AMP [¹²⁵I] kit "Du Pont-Daiichi", Daiichi Kagaku Yakuhin). An increase in cAMP production depending on 20 the concentration of the human PTH partial peptide (the 1-34 positions) added as a standard was always observed. hPTH obtained in Example 1 described above also had an activity approximately similar to this activity (refer to Fig. 11).

The hPTH gene provided by the present invention has the novel DNA sequence and shows a high expression rate of hPTH. Further, the present invention first made it possible to allow hPTH to express in 25 large amounts in the system using *E. coli* as the host. Furthermore, hPTH can be produced more efficiently by the recombinant DNA technique of the present invention using the synthetic gene corresponding to hPTH, which serves to produce hPTH as a therapeutic agent or to study the biological role of hPTH in vivo.

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Sequence Listing

5 SEQ ID NO:1:

LENGTH: 252

10 TYPE:nucleic acid

STRANDEDNESS:double

15 TOPOLOGY:linear

MOLECULE TYPE:other nucleic acid, synthesizing DNA

20 FEATURES:

FEATURES KEYS:Mat peptide

LOCATION:1 . . . 252

25 IDENTIFICATION METHOD:E

FEATURES KEYS:mutation

LOCATION:7, 8, 9, 12, 15, 19, 21, 33, 36, 43, 51, 58, 60, 82, 109, 111,

30 114, 117, 120, 121, 123, 129, 130, 132, 147, 150, 153, 156, 177, 196, 197,
198, 201, 204, 216, 219

35 IDENTIFICATION METHOD:S

SEQUENCE DESCRIPTION:

40 TCT GTG TCC GAG ATT CAG TTA ATG CAT AAC CTT GCC AAA CAT TTG AAC 48

Ser Val Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn

45 1 5 10 15

TCC ATG GAG CGT GTA GAA TGG CTG CGT AAG AAG TTG CAG GAT GTG CAC 96

Ser Met Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His

50 20 25 30

AAT TTT GTT GCC TTA GGT GCC CCA TTG GCT CCT CGT GAT CCT GGT TCC 144

Asn Phe Val Ala Leu Gly Ala Pro Leu Ala Pro Arg Asp Ala Gly Ser

55

35 40 45
5 CAA AGA CCA CGT AAA AAG GAA GAC AAT GTC TTA GTT GAG AGC CAT GAA 192
Gln Arg Pro Arg Lys Lys Glu Asp Asn Val Leu Val Glu Ser His Glu
10 50 55 60
AAA TCC CTA GGC GAG GCA GAC AAG GCC GAT GTG AAT GTA TTA ACT AAA 240
15 Lys Ser Leu Gly Glu Ala Asp Lys Ala Asp Val Asn Val Leu Thr Lys
65 70 75 80
GCT AAA TCC CAG 252
20 Ala Lys Ser Gln
84
25

SEQ ID NO:2:

30 LENGTH: 263
35 TYPE:nucleic acid
STRANDEDNESS:double
35 TOPOLOGY:linear
MOLECULE TYPE:other nucleic acid, synthesizing DNA
40 FEATURES:
FEATURES KEYS:CDS
45 LOCATION:2...262
IDENTIFICATION METHOD:E
50 FEATURES KEYS:Mat peptide
LOCATION:5...256
IDENTIFICATION METHOD:E
55 FEATURES KEYS:mutation

LOCATION:11, 12, 13, 16, 19, 23, 25, 37, 40, 47, 55, 62, 64, 86, 113, 11
5 5, 118, 121, 124, 125, 127, 133, 134, 136, 151, 154, 157, 160, 181, 200, 2
01, 202, 205, 207, 220, 223, 259, 261, 263

10 IDENTIFICATION METHOD:S

SEQUENCE DESCRIPTION:

15 TATGTCTGTG TCCGAGATTG AGTTAATGCC TAACCTTGGC AAACATTTGA ACTCCATGGA 60
20 GCGTGTAGAA TGGCTGCGTA AGAAGTTGCA GGATGTGCAC AATTTTGTG CCTTAGGTGC 120
25 CCCATTGGCT CCTCGTGATG CTGGTTCCCA AAGACCACGT AAAAAGGAAG ACAATGTCTT 180
AGTTGAGAGC CATGAAAAAT CCCTAGGCCA GGCAAGACAAG GCCGATGTGA ATGTATTAAC 240
TAAAGCTAAA TCCCAGTAAT GAC 263

25

SEQ ID NO:3:

30

LENGTH:265

35

TYPE:nucleic acid

STRANDEDNESS:double

40

TOPOLOGY:linear

ANTISENSE:yes

45

SEQUENCE DESCRIPTION:

50 GATGGCTGATT ACTGGATTT AGCTTTAGTT AATACATTCAT CATGGGCCTT GTCTGCCCTCG 60
CCTAGGGATT TTTCATGGCT CTCAACTAAG ACATTGTCTT CCTTTTACG TGGTCTTGG 120
45 GAACCAGCAT CACGAGGAGC CAATGGGCA CCTAAGGCAA CAAAATTGTG CACATCCTGC 180
AACTTCTTAC GCACCCATTTC TACACGGCTCC ATGGAGTTCA AATGTTGCC AACGTTATCC 240
55 ATTAACTGAA TCTCGGACAC AGACA 265

55

SEQ ID NO:4

LENGTH:30

5 TYPE:nucleic acid

STRANDEDNESS:single

10 TOPOLOGY:linear

MOLECULE TYPE:other nucleic acid, sythesizing DNA

15 FEATURES

FEATURES KEYS:CDS

LOCATION:2...30

20 IDENTIFICATION METHOD:E

SEQUENCE DESCRIPTION:

25 TATGTCTGTG TCCGAGATTG AGTTAATGCA 30

30 SEQ ID NO:5

LENGTH:34

35 TYPE:nucleic acid

STRANDEDNESS:single

TOPOLGY:linear

40 ANTISENSE:yes

MOLECULE TYPE:other nucleic acid, sythesizing DNA

45 FEATURES

FEATURES KEYS:CDS

LOCATION:1...34

50 IDENTIFICATION METHOD:E

SEQUENCE DESCRIPTION:

55 AGCTTATGCA TTAACTGAAT CTCGGACACA GACA 34

5 SEQ ID:6

10 LENGTH:45

15 TYPE:nucleic acid

20 STRANDEDNESS:single

25 TOPOLOGY:linear

30 MOLECULE TYPE:other nucleic acid, sythesizing DNA

35 FEATURES

40 FEATURES KEYS:CDS

45 LOCATION:1...45

50 IDENTIFICATION METHOD:E

55 FEATURE DESCRIPTION:

TAACCTTGGC AAACATTGA ACTCCATGGA GCGTGTAGAA TGGCT

45

60 SEQ ID:7

65 LENGTH:45

70 TYPE:nucleic acid

75 STRANDEDNESS:single

80 TOPOLOGY:linear

85 MOLECULE TYPE:other nucleic acid, sythesizing DNA

90 FEATURES

95 FEATURES KEYS:CDS

100 LOCATION:1...45

110 IDENTIFICATION METHOD:E

115 SEQUENCE DESCRIPTION:

TTACCCAGCC ATTCTACACG CTCCATGGAG TTCAAATGTT TGCCA

45

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SEQ ID:8

10

LENGTH:30

TYPE:nucleic acid

15

STRANDEDNESS:single

20

TOPOLOGY:linear

MOLECULE TYPE:other nucleic acid, sythesizing DNA

25

FEATURES

FEATURES KEYS:CDS

30

LOCATION:1...30

IDENTIFICATION METHOD:E

35

SEQUENCE DESCRIPTION:

GCCTAAGAAG TTGCAGGATG TGCACAATT

30

40

SEQ ID:9

LENGTH:30

45

TYPE:nucleic acid

STRANDEDNESS:single

TOPOLOGY:linear

50

ANTISENSE:yes

MOLECULE TYPE:other nucleic acid, sythesizing DNA

55

FEATURES

FEATURES KEYS:CDS

60

LOCATION:1...30

IDENTIFICATION METHOD:E

5

SEQUENCE DESCRIPTION:

GCAACAAAAT TGTGCACATC CTGCAACTTC

30

10

SEQ ID:10

15

LENGTH:46

TYPE:nucleic acid

20

STRANDEDNESS:single

TOPOLOGY:linear

MOLECULE TYPE:other nucleic acid, synthesizing DNA

25

FEATURES

FEATURES KEYS:CDS

30

LOCATION:1...46

IDENTIFICATION METHOD:E

SEQUENCE DESCRIPTION:

35

TGTTGCCTTA GGTGCCCAT TGGCTCCTCG TGATGCTGGT TCCCAA

46

40

SEQ ID:11

LENGTH:46

45

TYPE:nucleic acid

STRANDEDNESS:single

50

TOPOLOGY:linear

ANTISENSE:yes

MOLECULE TYPE:other nucleic acid, synthesizing DNA

55

FEATURES

FEATURES KEYS:CDS

5 LOCATION:1...46

IDENTIFICATION METHOD:E

10 SEQUENCE DESCRIPTION:

TGGTCTTGG GAACCAGCAT CACGAGGAGC CAATGGGCA CCTAAG

46

15 SEQ ID:12

LENGTH:41

20 TYPE:nucleic acid

STRANDEDNESS:single

25 TOPOLOGY:linear

MOLECULE TYPE:other nucleic acid, synthesizing DNA

30 FEATURES

FEATURES KEYS:CDS

LOCATION:1...41

35 IDENTIFICATION METHOD:E

SEQUENCE DESCRIPTION:

40 AGACCACGTA AAAAGGAAGA CAATGTCTTA GTTGAGAGCC A

41

45 SEQ ID:13

LENGTH:41

TYPE:nucleic acid

50 STRANDEDNESS:single

TOPOLOGY:linear

55 ANTISENSE:yes

5 MOLECULE TYPE:other nucleic acid, synthesizing DNA

FEATURES

FEATURES KEYS:CDS

10 LOCATION:1...41

IDENTIFICATION METHOD:E

15 SEQUENCE DESCRIPTION:

TTTCATGGC TCTCACTAA GACATTGTCT TCCTTTTAC G

41

20 SEQ ID:14

LENGTH:42

25 TYPE:nucleic acid

STRANDEDNESS:single

30 TOPOLOGY:linear

MOLECULE TYPE:other nucleic acid, synthesizing DNA

FEATURES

35 FEATURES KEYS:CDS

LOCATION:1...42

40 IDENTIFICATION METHOD:E

SEQUENCE DESCRIPTION:

45 TCAAAATCC CTAGGCGAGG CAGACAAGGC CGATGTGAAT GT

42

50 SEQ ID:15

LENGTH:42

TYPE:nucleic acid

55 STRANDEDNESS:single

5 TOPOLOGY: linear

ANTISENSE: yes

10 MOLECULE TYPE: other nucleic acid, synthesizing DNA

FEATURES

15 FEATURES KEYS: CDS

LOCATION: 1 .. 42

16 IDENTIFICATION METHOD: E

20 SEQUENCE DESCRIPTION:

25 GTTAATACAT TCACATCGGC CTTGTCTGCC TGGCCTAGGG AT

42

25 SEQ ID: 16

LENGTH: 29

30 TYPE: nucleic acid

STRANDEDNESS: single

35 TOPOLOGY: linear

MOLECULE TYPE: other nucleic acid, synthesizing DNA

FEATURES

40 FEATURES KEYS: CDS

LOCATION: 1 .. 28

45 IDENTIFICATION METHOD: E

SEQUENCE DESCRIPTION:

50 ATTAACTAAA GCTAAATCCC AGTAATGAG

29

55 SEQ ID: 17

LENGTH: 27

5 TYPE:nucleic acid

STRANDEDNESS:single

10 TOPOLOGY:linear

15 ANTISENSE:yes

MOLECULE TYPE:other nucleic acid, sythesizing DNA

15 FEATURES

FEATURES KEYS:CDS

20 LOCATION:6...27

IDENTIFICATION METHOD:E

25 SEQUENCE DESCRIPTION:

25 GATCCTCATT ACTGGGATT AGCTTTA

27

30 **Claims**

1. DNA containing a synthetic gene encoding the human parathyroid hormone represented by the following DNA sequence:

35 **TCTGTG TCCGAGATTG AGTAAATGCA TAACCTTGGC**
AAACATTGGA ACTCCATGGA GCGTGTAGAA TGGCTGCGTA
40 **AGAAGTTGCA GGATGTGCAC AATTTTGTG CCTTAGGTGC**
CCCATTGGCT CCTCGTGATG CTGGTTCCCA AAGACCACGT
AAAAAGGAAG ACAATGTCTT AGTTGAGAGC CATGAAAAAT
45 **CCCTAGGCAGA GGCAGACAAG GCCGATGTGA ATGTATTAAC**
TAAAGCTAAA TCCCCAG

50 2. A method for producing the DNA of claim 1, which comprises enzymatically ligating a plurality of oligodeoxynucleotides one to another to form the DNA and inserting the DNA into a vector if necessary.

55 3. A vector containing the DNA of claim 1, wherein the synthetic gene is inserted into a control region of an Escherichia coli T7 promoter.

4. A host cell transformed by the DNA claimed in claim 1 or claim 3.

5. A method for producing human parathyroid hormone which comprises cultivating the transformant claimed in claim 4, accumulating human parathyroid hormone in a culture medium, and collecting the same.

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Fig. 1

TCT. GTG. TCC. GAG. ATT. CAG. TTA. ATG. CAT. AAC. CTT. GGC. AAA. CAT. TTG. AAC. TCC. ATG. GAG
Ser-Val-Ser-Glu-Ile-Gln-Leu-Met-His-Asn-Leu-Gly-Lys-His-Leu-Asn-Ser-MET-Glu

CGT. GTA. GAA. TGG. CTG. CGT. AAG. AAG. TTG. CAG. GAT. GTG. CAC. AAT. TTT. GTT. GCC. TTA. GGT. GCC
Arg-Val-Glu-Trp-Leu-Arg-Lys-Lys-Leu-Gln-Asp-Val-His-Asn-Phe-Val-Ala-Leu-Gly-Ala

CCA. TTG. GCT. CCT. CGT. GAT. GCT. GGT. TCC. CAA. AGA. CCA. CGT. AAA. AAG. GAA. GAC. AAT. GTC. TTA
Pro-Leu-Ala-Pro-Arg-Asp-Ala-Gly-Ser-Gln-Arg-Pro-Arg-Lys-Lys-Glu-Asp-Asn-Val-Leu

GT. GAG. ACC. CAT. GAA. AAA. TCC. CTA. GGG. GAG. GCA. GAC. AAG. GCC. GAT. GTG. AAT. GTA. TTA. ACT
Val-Glu-Ser-His-Glu-Lys-Ser-Leu-Gly-Glu-Ala-Asp-Lys-Ala-Asp-Val-Asn-Val-Leu-Thr

AAA. GCT. AAA. TCC. CAG
Lys-Ala-Lys-Ser-Gln

Fig. 2

10	20	30	40	50	60
TATGCTGTG	TCCGAGATC	AGTTAATGCA	<u>TAACCTTGGC</u>	AAACATTGA	ACTCCATGGA
ACAGACAC	<u>AGGCTCTAAG</u>	TCATTACGT	<u>ATGGAAACCG</u>	TTTGTAACT	<u>TGAGGTACCT</u>
<u>Nde I</u>	<u>Hinf I</u>			<u>Nco I</u>	
70	80	90	100	110	120
GGGTGTAGAA	<u>TGGCTGGTA</u>	<u>AGAAAGTTGCA</u>	<u>GGATGTGCAC</u>	<u>AATTGTGTG</u>	<u>CCTTAGGTCC</u>
CGCACATCTT	ACCGACGGAT	<u>TCTCAACGT</u>	<u>CCTACACCTG</u>	<u>TTAAAACAAAC</u>	<u>QGAATCCACG</u>
			<u>HgiA I</u>		
130	140	150	160	170	180
CCCATTGGCT	CCTCGTGTAG	CTGGTTCCCA	<u>AGAGCACCACGT</u>	AAAAAGGAAG	ACAAATGTCTT
<u>GGGTAACCGA</u>	<u>GGAGGCACTAC</u>	<u>GACCAAGGGT</u>	<u>TTCCTGGTGC</u>	<u>TTTTTCCCTC</u>	<u>TGTTACAGAA</u>
<u>Bgl I</u>				<u>Dde I</u>	
190	200	210	220	230	240
AGTTGAGACC	<u>CATGAAAT</u>	<u>CCCTAGGCGA</u>	<u>GGCAGACAAAG</u>	<u>GGCGATGTGA</u>	<u>ATGTTAAAC</u>
<u>TCAACTCTCG</u>	<u>GTACTTTTA</u>	<u>GGGATCCGGCT</u>	<u>CCGTCTGTC</u>	<u>CGGCTACACT</u>	<u>TACATAATTQ</u>
			<u>Avr II</u>		
250	260	270			
TAAAGCTAA	TCCCAGTAAAT	GAC			
<u>ATTTCGATTT</u>	<u>AGGGTCATTA</u>	<u>CTCCTAG</u>			
<u>Alu I</u>			<u>BamH I</u>		

Fig. 3

#1 5' TATGTCTGGCCGAGATTCACTTAATGCA^{3'}
 #2 3' ACAGACACAGGCTCTAAGTCATTACGTTATTGGCA^{5'}

 #3 5' TAACCTTGGCAAACATTGAACACTCCATGGAGCGTGTAGAAATGGCT^{3'}
 #4 3' ACCGGTTGTAAACTTGAGGTTACCTCGCACATCTTACCGACGGCATT^{5'}

 #5 5' GCGTAAGAAGTTGCAGGATGTCACAAATT^{3'}
 #6 3' CTTCAAACGTCCTACACCGTGTAAACCAACG^{5'}

 #7 5' TGTTGCCTTAGGTGCCCATGGCTCCTCGTGAATGCTGGTTCCCAA^{3'}
 #8 3' GAATCCACGGGTAACCGAGGACTACGACCAAGGGTTCTGGT^{5'}

 #9 5' AGACCCACGTAAAAAGGAAGACAATGGTCTTAGTTGAGAGCCA^{3'}
 #10 3' GCATTTCCCTCTGTACAGAATCAAACCTCGGTACTTT^{5'}

 #11 5' TGAAAAATCCCTAGGCCAGACAAGGCCATGTGAATGT^{3'}
 #12 3' TAGGGATCCGCTCCGGTCTGGCTACACTAACATAATTG^{5'}

 #13 5' ATTAACCTAAAGCTAAATCCCAGTAATGAG^{3'}
 #14 3' ATTCGGATTAGGGTCAATTACTCCTAG^{5'}

Fig. 4-1

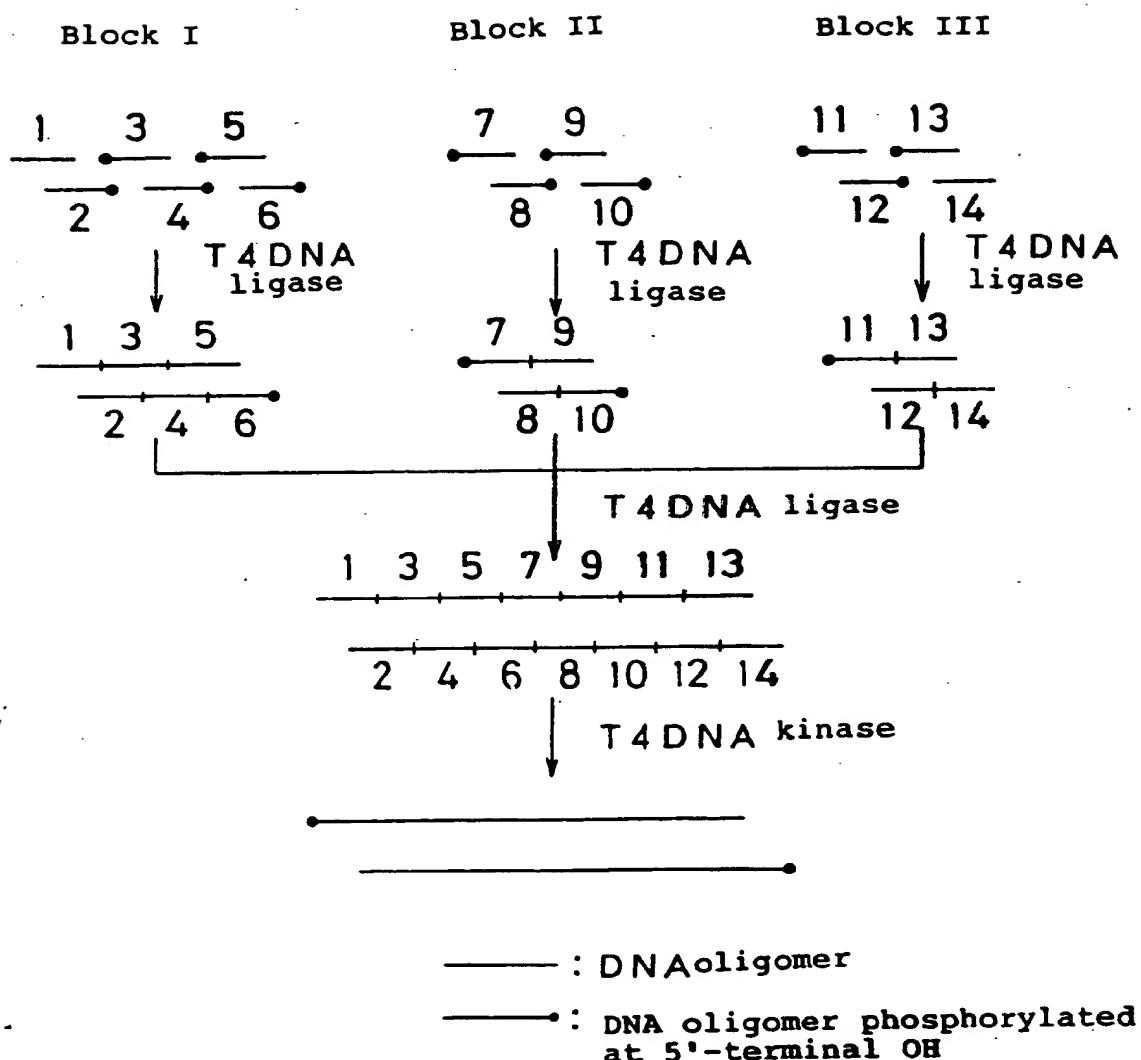


Fig. 4-2

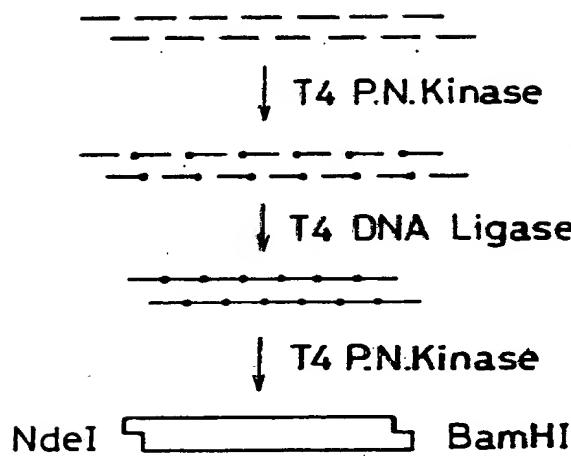


Fig. 5

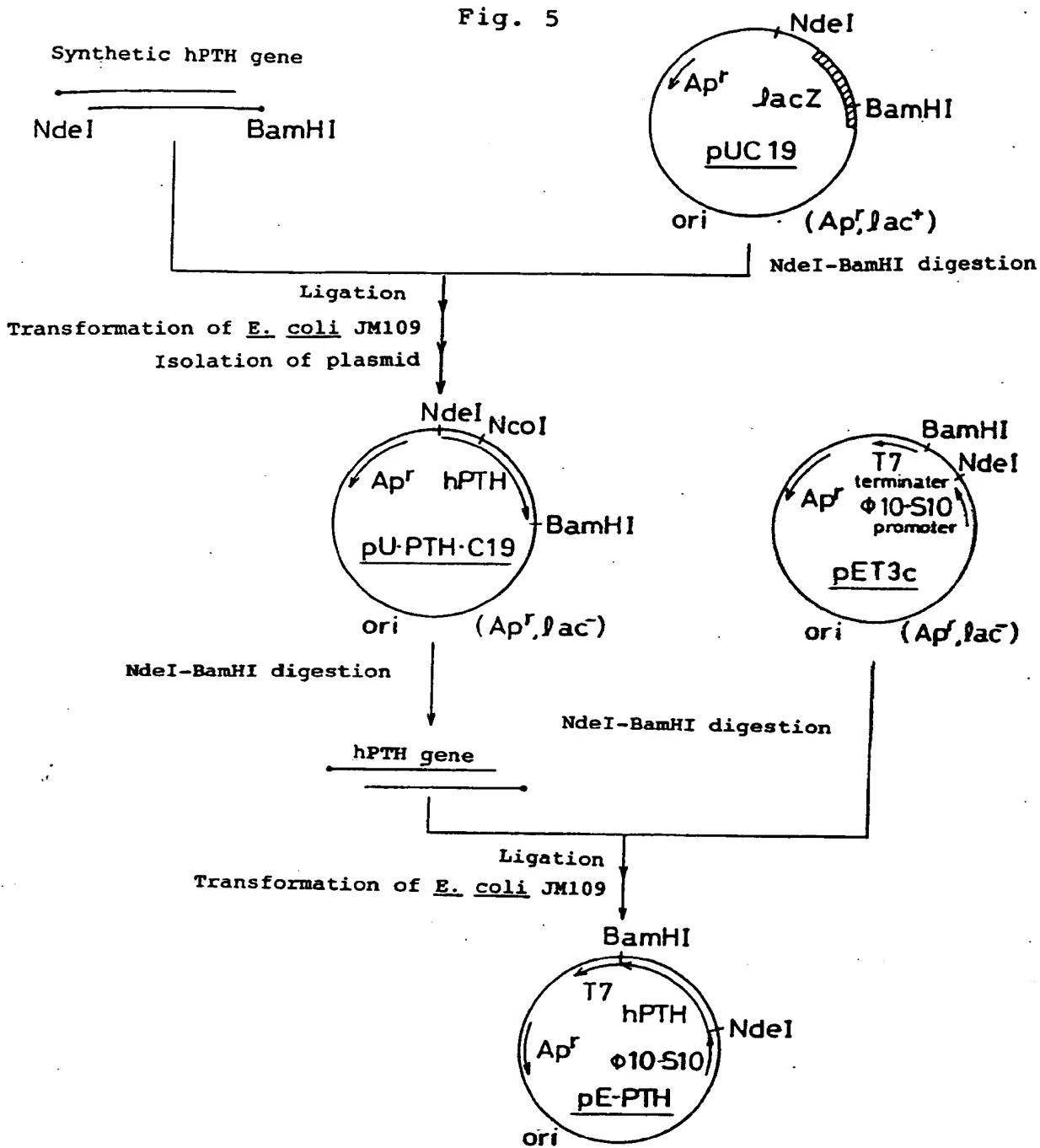


Fig. 6

1 2 3 4

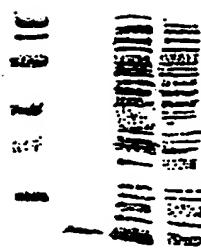


Fig. 7

1 2 3 4



Fig. 8

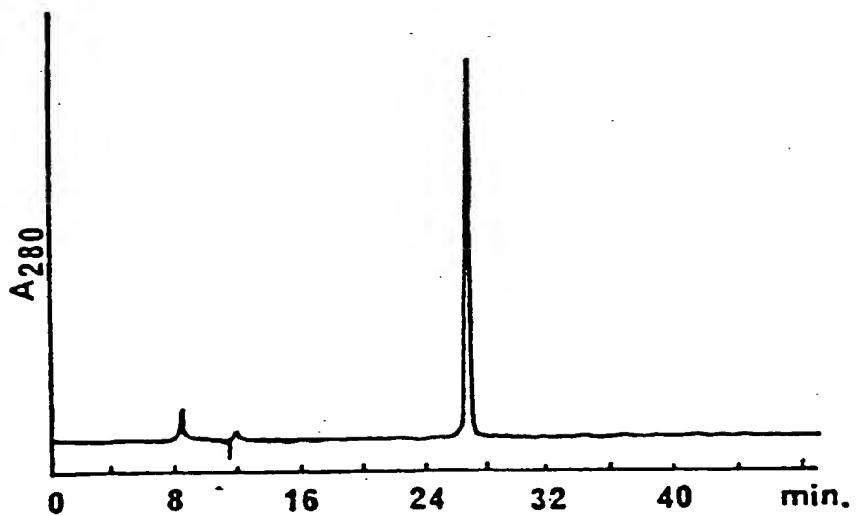


Fig. 9



Fig. 10

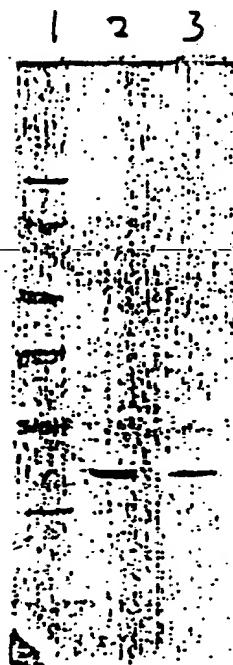
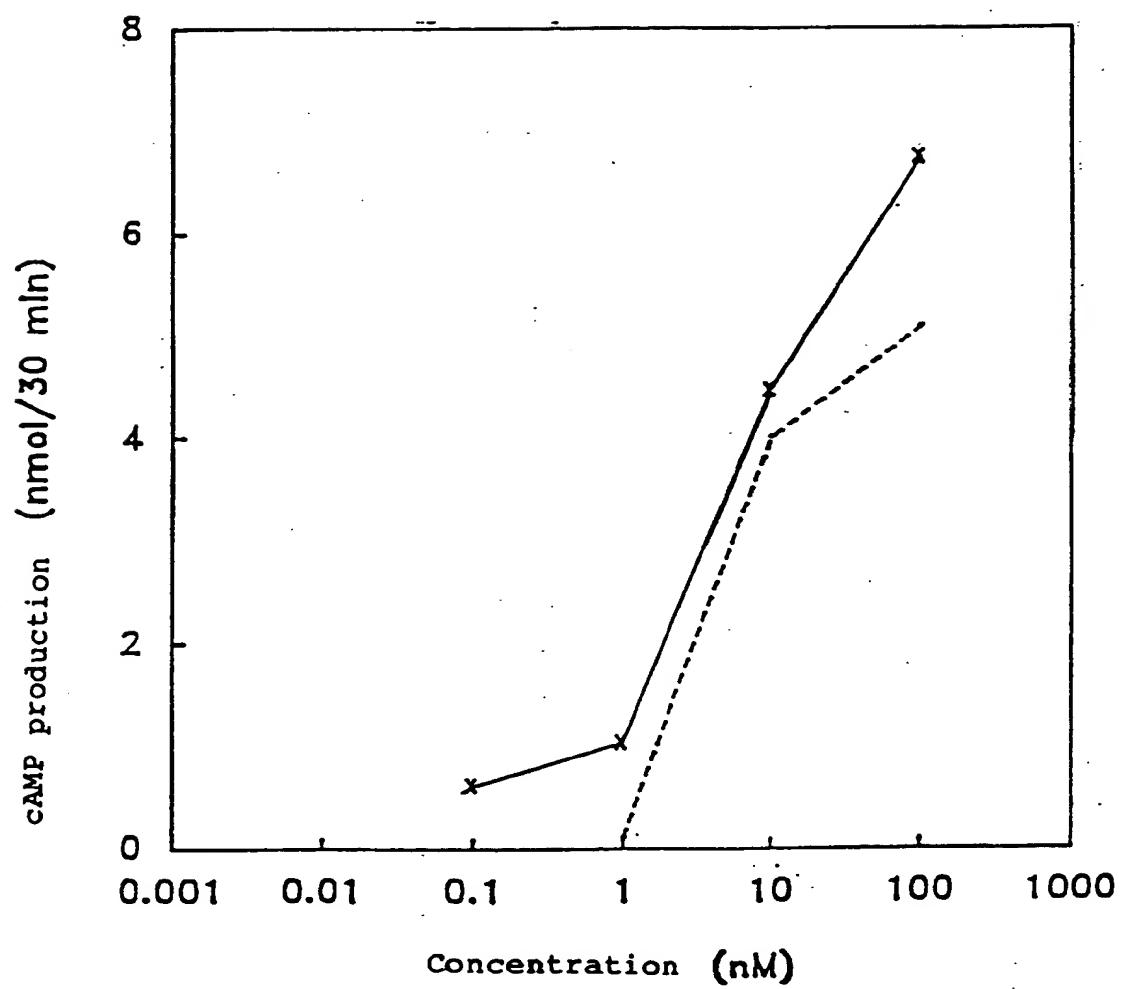


Fig. 11





European Patent
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EUROPEAN SEARCH REPORT

Application Number

EP 91 11 6281

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 5)
P, X	WO-A-9 105 050 (NATIONAL RESEARCH COUNCIL CANADA) * whole document * ---	1-5	C 12 N 15/16 C 07 K 15/00 C 12 P 21/02
D, Y	BIOCHEMISTRY AND CELL BIOLOGY vol. 64, no. 2, February 1986, pages 133-138, Canada; W.L. SUNG et al. "Hybrid gene synthesis: its application to the assembly of DNA sequences encoding the human parathyroid hormones and analogues" * whole article * ---	1-5	
D, Y	THE JOURNAL OF BIOLOGICAL CHEMISTRY vol. 263, no. 3, 25 January 1988, pages 1307-1313, Baltimore, US; S.A. RABBANI et al.: "Recombinant Human Parathyroid Hormone Synthesized in Escherichia coli" * whole article * ---	1-5	
A	GENE vol. 47, no. 2/3, 1986, pages 261-267, Amsterdam, NL; W.L. SUNG et al.: "Synthesis of mutant parathyroid hormone genes via site-specific recombination directed by crossover linkers" * whole article * ---	1-5	TECHNICAL FIELDS SEARCHED (Int. Cl.5) C 12 N 15/16 C 07 K 15/00 A 61 K 37/24
A	WO-A-8 803 165 (SELMER SANDE A.S.) * whole document * ---	1-5	
A	EP-A-0 178 863 (SHERING CORPORATION) * whole document * -----	3	
The present search report has been drawn up for all claims			
Place of search	Date of completion of the search		Examiner
BERLIN	05-12-1991		JULIA P.
CATEGORY OF CITED DOCUMENTS			
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